# Seed yield and physiological responses to deal with drought stress and late sowing date for promising lines of rapeseed (*Brassica napus* L.)

Maryam Teymoori<sup>1</sup><sup>(10)</sup>, Mohammad Reza Ardakani<sup>2</sup><sup>(10)</sup>\*, Amir Hossein Shirani Rad<sup>3</sup><sup>(10)</sup>, Mojtaba Alavifazel<sup>4</sup><sup>(10)</sup>, and Parisa Nejatkhah Manavi<sup>1</sup><sup>(10)</sup>

 <sup>1</sup>Department of Agronomy, North Tehran Branch, Islamic Azad University, Tehran, Iran, P.O. Box: 13919-37911
 <sup>2</sup>Department of Agronomy, Karaj Branch, Islamic Azad University, Karaj, Iran, P.O. Box: 31499-68111
 <sup>3</sup>Department of Oilseed Research, Seed and Plant Improvement Institute (SPII), Agricultural Research, Education and Extension Organization (AREEO), Iran, P.O. Box: 31359-33151
 <sup>4</sup>Department of Agronomy, Ahvaz Branch, Islamic Azad University, Ahvaz, Iran, P.O. Box: 61349-37333

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Abstract. The introduction of new genotypes of crop plants is among the most strategic research programmes, especially in arid and semi-arid regions. To study the effect of drought stress on seed yield and some physiological traits of promising lines of rapeseed at different sowing dates, an experiment was conducted for two years (2015-2017) in a semi-arid region of Iran. In this research, two conventional sowing dates were set in October 12 and November 1 (late sowing). Irrigation was carried out at two levels: normal irrigation (control) and irrigation interruption from the silique formation stage to the next stage (late-season drought stress). The genotypes included four promising lines (L1112, L1091, L1093, L1206), and a cultivar (Okapi) as a control. Results showed that delayed sowing and drought stress increased carbohydrate content and decreased seed yield, with the highest carbohydrate content and highest yield loss in L1112 and the lowest carbohydrate and lowest yield loss in the L1206 line. Among the physiological traits measured, stomatal resistance had the highest degree of correlation and the highest direct negative effect on seed yield, which declined with increasing stomatal resistance. L1112 had the highest stomatal resistance (52.76 s cm<sup>-1</sup>) in delayed sowing and drought stress conditions. Therefore, L1206 and L1112 were revealed to be resistant and sensitive lines, respectively.

K e y w ord s: *Brassica napus* L., carbohydrate, correlation, drought stress, seed yield, proline

#### INTRODUCTION

Due to the rainfall deficit in various regions of the world, which has caused drought, releasing the varieties/ lines of plants which are resistant to drought stress has a particular importance in the regions of the world with low levels of precipitation. The rapeseed (Brassica napus L.) crop contains 40-44% oil and is considered to be one of the most important edible oilseeds (Ashkiani et al., 2020; Cashin et al., 2014; FAO, 2018), being the third annual oil crop in the world after soybean and oil palm (Enjalbert et al., 2013). One of the most critical factors that inhibits the development of the area under cultivation and prevents the successful production of rapeseed in semi-arid regions is moisture deficiency at the end of the growth period (reproductive stage) to which it is very susceptible (Nasiri et al., 2017). Among the various abiotic stresses, drought stress is the most severe, destructive and significant restricting environmental factor for sustainable agricultural production (Ardakani et al., 2009; Davaran Hagh et al., 2016; Pace and Benincasa, 2010; Yuncai and Schmidhalter, 2005).

<sup>\*</sup>Corresponding author e-mail: mreza.ardakani@gmail.com

Hammac *et al.* (2017) assessed the effect of drought stress on seed yield, as well as the protein and fatty acids content of the rapeseed and reported that drought stress at the flowering stage had significant effects on unsaturated fatty acids, protein, and seed yield. Photosynthesis limitation through metabolic destruction is a complicated phenomenon, in which pigments have a protective task (Jaleel *et al.*, 2009; Khan *et al.* 2010). Chlorophyll loss under drought stress causes damages due to the formation of free oxygen radicals (Gaspar *et al.*, 2002; Hoseinzade *et al.*, 2016; Ilkaei *et al.*, 2011). Drought stress mediated by soil moisture deficiency in flowering and silique development stages decreased chlorophyll a and b contents in various rapeseed lines compared with the irrigation treatment (Din *et al.*, 2011; Grewal, 2010; Saranga *et al.*, 2001).

Due to the water deficiency problem in arid and semiarid regions across the globe such as Iran, the feasibility of late rapeseed and the simultaneous cultivation of the crop with spring crops may be considered as a management strategy for the optimal use of irrigation water. Additionally, the irrigation of rapeseed at the late growth season coincides with the first irrigations of spring crops after rapeseed. It is also very important to select superior promising lines that can potentially respond well to delayed cultivation and water deficiency problems, which was the aim of this study.

### MATERIALS AND METHODS

This experiment was conducted in the research farm of the Seed and Plant Improvement Institute (Karaj, Iran) during 2015-2017. The geographic coordinates of the experimental farm were 35°49' N and 51°6' E, with an altitude of 1321 m above sea level. This region has a warm and dry Mediterranean climate zone, as well as a dry moisture regime due to the cold and humid winters and warm and dry summers, there are from 150 up to 180 dry days per year based on meteorological information. The average regional yearly precipitation is 243 mm. Rainfall occurs mainly late in the fall and early in spring. The average maximum annual temperature is 28°C in July, and an average minimum temperature of 1°C occurs in January. The average 30-year soil temperature is 14.5°C. The amberothermic graph for the two years of the experiment is illustrated in Fig. 1.

The soil was sampled before fertilization from a depth of 0-30 cm. The soil had the following characteristics: organic carbon (0.64%), electrical conductivity (EC) of 1.70 ds m<sup>-1</sup>, soil nitrogen (0.06%), and absorbable phosphorus and potassium (10.3 and 275 mg kg<sup>-1</sup> of soil, respectively). The soil texture was clay loam (sand 28.80%, clay 41.88%, and silt 29.32%).

A factorial split-plot experiment was conducted in a completely randomized design with three replications over two years (2015-2017). The sowing dates were October 12 (conventional date) and November 1 (late sowing) with irrigation as the variable in the main plots, this took place at two levels: normal irrigation (control) and discontinuing irrigation from the silique forming stage (development of fruit stage, BBCH: 71) to the next stage (late-season drought stress). The subplots included promising new lines of open-pollinated rapeseed, that were L1112, L1091, L1093, L1206, which were originally released in Iran according to the national breeding programs for oilseeds crops and an open-pollinated cultivar, Okapi (French origin) as a control to be compared with the new lines. Each plot included six rows, with a length of 6 m, a distance of 30 cm between the lines, and a plant distance of 4 cm on the lines. The two outer rows were considered to be the border, and the four inner lines were used to evaluate various plant traits.

During both cropping years, fertilization was carried out by using nitrogen (urea 150 kg ha<sup>-1</sup>, one third was applied at sowing time, one third at the stem elongation stage and one third at the beginning of the flowering stage), phosphorus (100 kg ha<sup>-1</sup> Triple-Superphosphate at sowing time) and potassium (50 kg ha<sup>-1</sup> Potassium Sulphate at sowing time).



Fig. 1. Climatology curve (amberotermic graph) for Karaj province from October 2015 to July 2017.

Treflan herbicide was evenly sprayed at a volume of 2.5 l ha<sup>-1</sup>. The fertilizers and herbicide were incorporated into the soil with a light disc. Pests, in particular the mealy aphid, were controlled through the application of Metasystox pesticide (1.5 l ha<sup>-1</sup>). Irrigation was implemented using siphon pipes based on 80 mm evaporation from the surface of a Class A evaporation basin and water deficiency stress treatments consisted of eight common irrigation steps, of which only six steps were carried out and irrigation cessation took place from the silique formation stage to the next stage and the amount of water used in these treatments was 5120 and 3840 m<sup>3</sup> ha<sup>-1</sup>, respectively (with due consideration of precipitation during the growth period for both years). Ten plants were randomly selected from each experimental plot to determine the plant traits. The relevant physiological traits were evaluated through the random selection of the youngest developed leaves of the third internode from the top of the plant per experimental plots during the fruit development stage (BBCH: 75). In order to measure the leaf relative water content (LRWC) 10 leaf samples were randomly selected from each experimental plot and this parameter was calculated according to Ferrat and Loval (1999) using Eq. (1):

$$LRWC(\%) = \frac{FW - DW}{TW - DW} 100,$$
(1)

where: *FW*, *DW*, and *TW* denote leaf fresh weight, dry weight, and the weight of inflammation, respectively.

Stomatal resistance (SR) was determined using a porometer device (Automatic Porometer AP4). Canopy temperature (CT) was measured using an infrared thermometer with an emission coefficient of 0.99. In order to measure the leaf proline content (LPC), 20 well-developed leaves were randomly selected from each experimental plot, and the amount of LPC was measured according to Bates *et al.* (1973). Carbohydrate of leaf solution (CLS) was calculated by randomly selecting 20 well-developed leaves from each experimental plot according to Dubois (Dubois *et al.*, 1956). Chlorophyll content (CC) was determined based on Arnon (1949) using Eq. (2):

# CC: (0.020) (OD 645) - (0.008) (OD 663) V/W, (2)

where: CC represents the total chlorophyll content (mg g<sup>-1</sup> of leaf fresh weight), OD denotes the absorbance of light at corresponding wavelengths, V shows the volume (ml) of acetone 80%, and W indicates leaf fresh weight. In order to measure seed yield, the margin and 0.5 m were removed from the top and bottom of the sowing rows, plants from the four middle rows of each plot were surface-cut in an area of 6 m<sup>2</sup>.

Compound data analysis covering the two-year experiment was completed using SAS software (Ver. 9). Excel software was used to plot the graphs. The mean values were compared using Duncan's multiple range test at a 5% probability level. Correlation and stepwise regression were analysed with SAS software (Ver. 9), and a path analysis was implemented using PATH software.

# RESULTS AND DISCUSSION

The results of the ANOVA for *LRWC* showed the significance of the main effects of the year, sowing date, irrigation, and lines. Also, the interactions between the year, sowing date, and irrigation, as well as the interactions of the year, planting date, and lines were significant with regard to the *LRWC* traits (p < 0.01) (Table 1).

The interactions between the year, sowing date, and irrigation showed that delayed sowing and drought stress significantly reduced the LRWC for both of the experimental years. The lowest LRWC (78.21%) occurred on the sowing date of November 1 with irrigation interruption from the silique formation stage in the first year, and the highest LRWC (93.74%) was achieved on the sowing date of October 12 and with normal irrigation (Table 2). In this experiment, irrigation enhanced water absorption by the rapeseed roots due to raising the available moisture to the roots, thereby raising the *LRWC*. On the other hand, water elimination as a result of the silique formation stage decreased LRWC. Also, precipitation undergoes high oscillations in the Karaj meteorological data; hence, it is probable that some of the important growth stages of the rapeseed crop were affected by reduced soil water potential due to water deficiency in this experiment. Genetically, drought resistance is correlated with LRWC and osmotic regulation (Cardoso et al., 2019; Reynolds et al., 2016). Khan et al. (2010) declared that LRWC was reduced in 5 rapeseed cultivars including Con-III, Hyola, Shiralee, Rainbow and two mutants of Rainbow cultivar under drought stress at the flowering, siliquae formation and maturity stages. The LRWC was uppermost (84.05%) for L1206 (Table 3). It seems that L1206 was more compatible with the climatic condition of this study, but L1112 had less compatibility with the late sowing date and climatic condition of the presented research due to the lower LRWC in the same situation. On the other hand, the LRWC is a physiological trait that has frequently been proposed as a selection criterion for drought tolerance in rapeseed (Wu et al., 2018).

Analysis of variances in this study showed that SR was influenced significantly by the sowing date and irrigation treatments. Furthermore, significant differences were observed between the two years and among the studied lines (p < 0.01) (Table 1). The interaction of the treatments was significant with regard to SR (0.01 ) (Table 1) and, thus, mean values were compared according to the interactions of year, sowing date, irrigation treatment, and lines (Fig. 2). Results showed that drought stress in general increased SR significantly (Fig. 2). Photosynthesis

**Table 1.** ANOVA for leaf relative water content (*LRWC*), stomatal resistance (SR), canopy temperature (CT), chlorophyll content (CC), leaf proline content (LPC), carbohydrate of leaf solution (CLS), and seed yield of the studied lines (2015-2017)

	df -	M.S.						
S.U.V.		LRWC	SR	СТ	LPC	CLS	CC	Seed yield
Year	1	256.6 **	46.99 **	29.12 **	199.1 **	395.5 **	0.138	$6.68 \times 10^6 **$
Error	4	0.18	0.97	0.43	1.006	0.05	0.027	$5.12 \times 10^4$
Sowing date (a)	1	2533 **	3852 **	1339 **	1364 **	7146 **	11.54 **	$1.06 \times 10^7 **$
Year $\times$ a	1	1.02 **	19.80 **	12.31 **	2.257 **	55.12 **	0.003	$8.55  imes 10^4$
Irrigation (b)	1	755.3 **	905.4 **	365.3 **	274.8 **	1647 **	2.594 **	$4.32 \times 10^7 **$
Year $\times$ b	1	2.84 **	0.22	3.38 **	0.077	10.28 **	0.004	$9.21  imes 10^4$
$\mathbf{a} \times \mathbf{b}$	1	28.80 **	62.05 **	0.81	13.60 **	4.04 **	0.070 *	$8.35 \times 10^5 *$
Year $\times$ a $\times$ b	1	76.27 **	1.02 *	2.70 **	37.92 **	28.69 **	0.007	$6.32 \times 10^5$
Error	12	0.04	0.21	0.23	0.169	0.28	0.008	$1.41 \times 10^{5}$
Line (c)	4	14.94 **	16.48 **	5.60 **	6.562 **	26.81 **	0.036 **	$2.01 \times 10^6 **$
$Year \times c$	4	10.36 **	13.80 **	5.19 **	5.22 **	28.29 **	0.041 **	$2.10 \times 10^{6}$ **
$a \times c$	4	5.13 **	6.61 **	1.18 *	2.68 **	6.91 **	0.010	$4.00 \times 10^{5}$
$Year \times a \times c$	4	7.34 **	5.51 **	2.25 **	3.177 **	8.97 **	0.010	$2.96 \times 10^5$
$b \times c$	4	0.21	0.16	0.07	0.063	1.04 *	0.003	$5.40  imes 10^4$
$Year \times b \times c$	4	0.04	0.28	0.14	0.160	1.65 **	0.003	$2.12 \times 10^4$
$a\times b\times c$	4	0.10	0.53	0.06	0.051	0.90	0.005	$1.50  imes 10^4$
$Year \times a \times b \times c$	4	0.30	0.79 *	0.09	0.041	0.80	0.002	$1.10 \times 10^4$
Error	64	0.12	0.22	0.40	0.31	0.37	0.005	$4.58 \times 10^{5}$
C.V. (%)		0.41	2.37	2.31	3.47	1.55	5.42	19.1

Significant at \*0.01 , <math>\*\*p < 0.01.

reduction under drought stress has been reported by other researchers, this phenomenon is associated with non-stomatal factors (Papanatsiou *et al.*, 2019; Trueba *et al.*, 2019).

A comparison of the rapeseed crop responses in the two years of the study revealed that the SR level was lower in the first year while it increased significantly in the second year (Fig. 2). Considering the results, it seems that inappropriate environmental conditions, such as an undesirable temperature and unequal distribution of precipitation in the second year, have brought about increased SR due to the negative effect on rapeseed internal metabolism. On the other hand, the lower SR in the first year are a result in part of the optimal climate conditions and precipitation in the first year (Fig. 1).

In almost all of the test plots, late sowing significantly amplified the SR (Fig. 2). With the November 1 sowing date, rapeseed could desirably stabilize the status of the stomatal cells since it had a shorter growth period and encountered fewer unsuitable environmental conditions than on October 12 (Henry et al., 2019). The mechanisms (such as stomatal closure and leaf area decrease) that bring about resistance to drought through diminishing water losses usually induce the reduction of carbon dioxide absorption. Osmotic pressure increases resistance to drought through the maintenance of crop turgor, but the enhancement of soluble substance concentrations, besides energy consumption, might have an undesirable effect on osmotic regulation. The highest SR value (52.76 s cm<sup>-1</sup>) was recorded in the second year and on the November 1 sowing date in L1112 under drought stress (irrigation interruption from the silique formation stage to the next stage). L1206 had the lowest SR (49.43 s cm<sup>-1</sup>) among the other examined lines in the unfavourable second year experiment with delayed cultivation (November 1) and drought stress (irrigation interruption from the silique formation stage). With open stomata under adverse conditions and

Treatments			L DIVG		LPC	CLS
Year	Sowing date	Irrigation	(%)	(°C)	(µmol g <sup>-1</sup> fresh weight)	(mg g <sup>-1</sup> fresh weight)
	0 ( 1 10	Control	92.91 <sup>b</sup>	24.82 <sup>g</sup>	20.68 <sup>b</sup>	25.85 <sup>h</sup>
2015- 2016 November 1	Drought stress	85.01 °	$27.84^{\text{ f}}$	15.90 <sup>d</sup>	$32.07^{\text{ f}}$	
	NT 1 1	Control	80.96 <sup>f</sup>	30.73 <sup>d</sup>	11.86 <sup>f</sup>	42.04 <sup>d</sup>
	November 1	Drought stress	78.21 <sup>h</sup>	34.01 <sup>b</sup>	10.68 <sup>g</sup>	49.47 <sup>b</sup>
	0.4.1	Control	93.74 <sup>a</sup>	24.53 <sup>g</sup>	21.90 <sup>a</sup>	29.28 <sup>g</sup>
2016- 2017 Nover	October 12	Drought stress	89.65 °	28.82 °	19.28 °	38.62 °
	NT 1 1	Control	85.35 <sup>d</sup>	32.32 °	15.89 <sup>d</sup>	44.71 °
	November 1	Drought stress	80.03 <sup>g</sup>	35.68 <sup>a</sup>	12.36 °	51.35 <sup>a</sup>

**Table 2.** Interactions of year, sowing date, and irrigation on leaf relative water content (*LRWC*), canopy temperature (CT), leaf proline content (LPC), and carbohydrate of leaf solution (CLS) of the studied lines (2015-2017)

Means in a column and a treatment followed by the same letter are not significantly different at 5% level.

**Table 3.** Interactions of year, sowing date, and line on leaf relative water content (*LRWC*), canopy temperature (CT), leaf proline content (LPC), and carbohydrate of leaf solution (CLS) of the studied lines (2015-2017)

Treatments				СТ	LPC	CLS
Year	Sowing date	Line	(%)	(°C)	(µmol g⁻¹ fresh weight)	(mg g <sup>-1</sup> fresh weight)
		L1112	87.60 <sup>g</sup>	26.85 <sup>i</sup>	17.72 <sup>f</sup>	29.96 <sup>1</sup>
		L1091	91.33 °	25.25 <sup>k</sup>	19.73 <sup>de</sup>	26.89 <sup>n</sup>
	October 12	L1093	86.91 <sup>h</sup>	27.32 <sup>hi</sup>	16.62 <sup>g</sup>	31.34 <sup>k</sup>
		L1206	90.72 <sup>de</sup>	25.70 <sup>k</sup>	19.44 <sup>e</sup>	27.51 <sup>n</sup>
		Okapi	88.23 <sup>f</sup>	26.53 <sup>ij</sup>	17.93 <sup>f</sup>	29.12 <sup>m</sup>
2015-2016						
		L1112	79.59 °	32.48 <sup>ef</sup>	11.23 <sup>kl</sup>	45.83 °
		L1091	80.26 <sup>n</sup>	31.68 <sup>g</sup>	11.69 <sup>k</sup>	43.72 <sup>g</sup>
	November 1	L1093	78.96 <sup>p</sup>	33.07 <sup>de</sup>	10.93 <sup>1</sup>	47.65 <sup>cd</sup>
		L1206	79.17 <sup>p</sup>	32.80 de	11.00 <sup>kl</sup>	47.11 <sup>d</sup>
		Okapi	79.95 <sup>no</sup>	31.82 fg	11.52 <sup>kl</sup>	44.46 <sup>f</sup>
		L1112	90.56 °	27.72 <sup>h</sup>	19.66 de	36.50 <sup>h</sup>
		L1091	91.03 <sup>cd</sup>	27.13 <sup>hi</sup>	20.25 <sup>cd</sup>	34.76 <sup>i</sup>
	October 12	L1093	91.43 °	26.89 <sup>i</sup>	20.52 bc	34.38 <sup>i</sup>
		L1206	92.99 ª	25.69 <sup>k</sup>	21.38 <sup>a</sup>	31.78 <sup>jk</sup>
		Okapi	92.47 <sup>b</sup>	25.94 <sup>jk</sup>	21.15 <sup>ab</sup>	32.33 <sup>j</sup>
2016-2017						
		L1112	81.45 <sup>m</sup>	34.97 <sup>a</sup>	13.13 <sup>j</sup>	49.74 <sup>a</sup>
		L1091	82.59 <sup>k</sup>	33.97 <sup>bc</sup>	14.20 <sup>i</sup>	48.21 °
	November 1	L1093	83.48 <sup>j</sup>	33.39 <sup>cd</sup>	14.83 <sup>hi</sup>	46.99 <sup>d</sup>
		L1206	84.05 <sup>i</sup>	33.14 de	15.01 <sup>h</sup>	46.20 °
		Okapi	81.89 <sup>1</sup>	34.51 <sup>ab</sup>	13.44 <sup>j</sup>	49.01 <sup>b</sup>

Means in a column and a treatment followed by the same letter are not significantly different at 5% level.



Fig. 2. Interactions of year, sowing date, irrigation, and line on stomatal resistance (SR) of the studied lines (2015-2017). Means with the same letter are not significantly different.

drought stress, the line received further  $CO_2$  and continued photosynthetic activity, therefore, L1206 had a superior performance in such conditions. However, L1112 had the highest SR with faster closure of stomata and reduced photosynthesis under adverse conditions (delayed planting date and drought stress). Research on three pea genotypes suggested that water deficiency caused a significant increase in SR (Krouma, 2010). Hence, drought stress and delayed sowing resulted in a rise in SR, and more resistant lines presented lower SR values.

The main effects of the year, sowing date, and irrigation were significant with regard to CT and also there were significant differences between the lines. The interaction of the effect of year, sowing date and irrigation was significant on CT (p < 0.01) (Table 1). The results of mean comparisons in the interaction effect of the year, sowing date and irrigation showed that the second year had less favourable weather conditions (Fig. 1) and resulted in an increase in CT (35.68°C) with delayed sowing (November 1) and drought stress (irrigation interruption from the silique formation stage). CT was lowermost with the October 12 sowing date and normal irrigation in both experimental years (Table 2). Delayed sowing with drought stress probably resulted in more rapid stomatal closure and lower levels of gas exchange, eventually increasing CT in the plant.

The results showed that the interaction of the sowing date for the two years and lines had a significant effect on CT (p < 0.01) (Table 1). A comparison of mean values revealed that the CT of the studied lines increased significantly through late sowing in both years of the experiment and that the highest CT value was observed in the second year for late sowing in L1112 (34.97°C), but L1206 (33.14°C) had the lowest CT in the same situation (Table 3). For the first sowing date, the crop was able to regulate its internal reactions in a way that CT did not increase to any significant extent. Increasing CT for the delayed sowing date is indicative of inappropriate environmental

conditions for optimal crop growth. L1112 revealed a lower compatibility with the conditions of the experiment due to a relatively higher CT and, in comparison with the other lines, was not able to desirably overcome the environmental conditions. Araus *et al.* (2008) and Balota *et al.* (2007) believe that lines with lower CT are more appropriate for planting in arid regions and would have increased resistance in these conditions.

The results of ANOVA showed that the main effects of year, sowing date, and irrigation treatments were significant for LPC and that there were notable differences between lines. LPC was also significantly influenced by the interactions of year, sowing date, and irrigation, as well as by interactions of year, sowing date, and lines (p < 0.01) (Table 1).

The mean comparison revealed that LPC was affected by the interaction of the sowing date and irrigation treatments in both years of the study (Table 2). As shown in Table 2, delayed sowing and drought stress decreased the LPC. Rapeseed lines may have used a pathway other than the LPC elevation pathway to cope with the stress they were subjected to because LPC production is, in turn, an energy-intensive pathway for the plant and this would reduce optimal plant yield. In stressful situations, the plant first tries to use less energy-intensive methods to cope. The highest LPC (21.90 µmol g<sup>-1</sup> leaf fresh weight) was obtained in the second year, October 12 sowing date, and normal irrigation, and the lowest level (10.68  $\mu$ mol g<sup>-1</sup> leaf fresh weight) was observed in the first year, November 1 sowing date, and drought stress. Drought stress leads to stomatal closure and the accumulation of LPC (Drake et al., 2013). Sanchez-Rodriguez (2010), however, reported a decreased amount of LPC in a stress-resistant cultivar of tomatoes. The interaction of the year and sowing date on different lines showed that the amount of LPC was significantly higher in the second year for most of the lines planted when the two years are compared, and late sowing

caused a significant decrease in LPC (Table 3). As mentioned above, it seems that the studied lines did not use an energy-intensive proline pathway to cope with stress conditions. However, L1206 had a higher LPC (15.01  $\mu$ mol g<sup>-1</sup> leaf fresh weight) than the other lines for the delayed sowing date of November 1 and in the second year. LPC acts as an osmotolerant factor and as a nutritional source (Chun *et al.*, 2018).

The ANOVA for CLS in the silique development stage showed that this parameter was significantly influenced by the sowing date and irrigation treatments. There were significant differences between the two years of the experiment and also between the lines. Interactions of the year, sowing date and irrigation; interactions of the year, sowing date and lines; interactions of the year, irrigation, and lines also had a significant influence on CLS content (p < 0.01) (Table 1). Concerning the climate condition in the first year of this research, precipitation dispersion was desirable early in the growth season, and the elevated vegetative growth led to diminish CLS.

The interactions of the year, sowing date and irrigation treatments showed that the highest amount of CLS (51.35 mg g<sup>-1</sup> leaf fresh weight) were observed in the second year, late sowing (November 1), and drought stress treatments (Table 2). By creating unfavourable conditions for the normal activities of the plant, drought stress and delayed sowing enable the plants to cope with these adverse conditions, hence, increasing CLS is one of the coping strategies (Yan *et al.*, 2016).

The interactions between the year, planting date, and lines showed that the highest amount of CLS (49.74 mg g<sup>-1</sup> leaf fresh weight) obtained in L1112 on the November 1 sowing date in the second year, indicated the higher susceptibility of this line to delayed sowing. The lowest level of CLS (46.20 mg g<sup>-1</sup> leaf fresh weight) was observed in L1206 (Table 3). A comparison of the mean interactions between year, irrigation and lines also showed that L1112 contained the highest amount of CLS (46.44 mg g<sup>-1</sup> leaf fresh weight) with delayed sowing and irrigation interruption from the silique formation stage in the second year. In the same conditions (delayed sowing and drought stress in the second year), the lowest level of CLS (43.26 mg g<sup>-1</sup> leaf fresh weight) was observed in L1206 (Table 4).

As mentioned in the LPC section, the production of LPC is very energy intensive for the plant to cope with adverse conditions. Therefore, the presented lines used less energy-intensive osmoregulation and increased CLS strategies to cope with the unfavourable conditions of delayed sowing and drought stress. Among the investigated lines, L1112 was able to cope with these conditions through a further synthesis of CLS due to its higher sensitivity to experimental situations.

 

 Table 4. Interactions of year, irrigation, and line on carbohydrate of leaf solution (CLS) of the studied lines (2015-2017)

2015-2016         L1112         34.34 i           L1091         32.02 k           L1093         35.90 h           L1206         34.24 i           Okapi         33.23 j           L1112         41.45 d           L1091         38.60 f           L1093         43.08 c           L1093         43.08 c           L1093         43.08 c           L1093         43.08 c           L1206         40.38 c           Okapi         40.35 c           L1112         39.81 c           L1112         39.81 c           L1091         37.58 g           Control         L1093         36.53 h           L1206         34.72 i           Okapi         36.33 h           L112         46.44 a           L1091         35.39 b           L1091         45.39 b           L1093         44.84 b           L1093         44.84 b           L1093         44.84 b           L1206         43.26 c           Okapi         45.01 b	Year	Irrigation	Line	CLS (mg g <sup>-1</sup> fresh weight)
2015-2016         L1091         32.02 k           2015-2016         L1206         34.24 i           Drought stress         L1112         41.45 d           L1091         38.60 f         11091           L1093         43.08 c         11091           L1093         43.08 c         11206           L1091         38.60 f         11093           L1091         37.58 g         11206           Control         L1091         37.58 g           L1091         37.58 g         11091           Control         L1093         36.53 h           L1206         34.72 i         00kapi           Okapi         36.33 h         1112           L1091         35.39 b         1112           Drought stress         L1091         36.53 h           L1112         46.44 a         11091           L1093         36.33 b         11093           L1091         45.39 b         11093           L1093         44.84 b         11206           L1093         44.84 b         1206           L1206         43.26 c         00kapi           Okapi         45.01 b         50.1 b			L1112	34.34 <sup>i</sup>
2015-2016         Control         L1093         35.90 h           2015-2016         L1206         34.24 i           Okapi         33.23 j           L1112         41.45 d           L1091         38.60 f           L1093         43.08 c           L1206         40.38 e           Okapi         40.35 e           L1091         37.58 g           Control         L1091           1112         39.81 c           L1091         37.58 g           Control         L1093           Ad.72 i         Okapi           Okapi         36.33 h           L1112         46.44 a           L1091         45.39 b           L1093         44.84 b           L1093         43.26 c           Okapi         43.26 c           Okapi         43.26 c           Okapi         45.01 b			L1091	32.02 <sup>k</sup>
2015-2016       L1206       34.24 i         Okapi       33.23 j         L1112       41.45 d         L1091       38.60 f         L1093       43.08 c         L1206       40.38 c         L1206       40.38 c         L1206       40.35 c         L1112       39.81 c         L1112       39.81 c         L1112       39.81 c         L1112       39.81 c         L1091       37.58 g         Control       L1093       36.53 h         L1206       34.72 i         Okapi       36.33 h         L1112       46.44 a         L1091       45.39 b         L1093       44.84 b         L1093       43.26 c         Okapi       43.26 c         Okapi       45.01 b		Control	L1093	35.90 <sup> h</sup>
$\begin{array}{cccc} & & & & & & & & & & & & & & & & & $			L1206	34.24 <sup>i</sup>
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2015 2016		Okapi	33.23 <sup>j</sup>
$\begin{array}{cccc} & L1091 & 38.60 & {}^{\rm f} \\ & L1093 & 43.08 & {}^{\rm c} \\ & L1206 & 40.38 & {}^{\rm e} \\ & U1206 & 40.33 & {}^{\rm e} \\ & U1206 & 40.35 & {}^{\rm e} \\ & U1112 & 39.81 & {}^{\rm e} \\ & L1091 & 37.58 & {}^{\rm g} \\ & L1091 & 37.58 & {}^{\rm g} \\ & L1091 & 36.53 & {}^{\rm h} \\ & L1206 & 34.72 & {}^{\rm i} \\ & 0{\rm kapi} & 36.33 & {}^{\rm h} \\ & L1112 & 46.44 & {}^{\rm a} \\ & L1091 & 45.39 & {}^{\rm b} \\ & L1093 & 44.84 & {}^{\rm b} \\ & L1206 & 43.26 & {}^{\rm c} \\ & 0{\rm kapi} & 45.01 & {}^{\rm b} \end{array}$	2015-2016		L1112	41.45 <sup>d</sup>
Drought stress         L1093         43.08 °           L1206         40.38 °           Okapi         40.35 °           L1112         39.81 °           L1091         37.58 °           L1091         37.58 °           Control         L1093           L1206         34.72 °           Okapi         36.53 h           L1206         34.72 °           Okapi         36.33 h           L1112         46.44 °           L1091         45.39 b           L1093         44.84 b           L1206         43.26 °           Okapi         45.01 b			L1091	38.60 <sup>f</sup>
2016-2017       L1206       40.38 °         0kapi       40.35 °         L1112       39.81 °         L1091       37.58 °         L1091       37.58 °         L1206       34.72 °         Okapi       36.33 °         L1112       46.44 °         L1091       45.39 °         L1093       44.84 °         L1206       43.26 °         Okapi       45.01 °		Drought stress	L1093	43.08 °
$\begin{array}{cccc} & & & & & & & & & & & & & & & & & $			L1206	40.38 <sup>e</sup>
$\begin{array}{cccc} & L1112 & 39.81 \\ & L1091 & 37.58 \\ & Control & L1093 & 36.53 \\ & L1206 & 34.72 \\ & 0 \\ & & 0 \\ & & 1112 & 46.44 \\ & & \\ & & L1091 & 45.39 \\ & & \\ & & L1093 & 44.84 \\ & & \\ & & L1206 & 43.26 \\ & & \\ & & 0 \\ & & & 0 \\ & & & & & \\ & & & &$			Okapi	40.35 <sup>e</sup>
$\begin{array}{cccc} & L1091 & 37.58 \\ & Control & L1093 & 36.53 \\ & L1206 & 34.72 \\ & 0 \\ & 0 \\ & 0 \\ & 2016-2017 & \\ & 0 \\ & 0 \\ & 1112 & 46.44 \\ & \\ & L1091 & 45.39 \\ & \\ & L1093 & 44.84 \\ & \\ & L1206 & 43.26 \\ & \\ & 0 \\ & \\ & 0 \\ & \\ & 0 \\ & \\ & \\$			L1112	39.81 <sup>e</sup>
$\begin{array}{cccc} \text{Control} & \text{L1093} & 36.53 & ^{\text{h}} \\ & \text{L1206} & 34.72 & ^{\text{i}} \\ \text{Okapi} & 36.33 & ^{\text{h}} \\ & \text{L1112} & 46.44 & ^{\text{a}} \\ & \text{L1091} & 45.39 & ^{\text{b}} \\ & \text{L1093} & 44.84 & ^{\text{b}} \\ & \text{L1206} & 43.26 & ^{\text{c}} \\ & \text{Okapi} & 45.01 & ^{\text{b}} \end{array}$			L1091	37.58 <sup>g</sup>
2016-2017 L1206 34.72 <sup>i</sup> Okapi 36.33 <sup>h</sup> L1112 46.44 <sup>a</sup> L1091 45.39 <sup>b</sup> L1093 44.84 <sup>b</sup> L1206 43.26 <sup>c</sup> Okapi 45.01 <sup>b</sup>		Control	L1093	36.53 <sup>h</sup>
2016-2017         Okapi         36.33 h           L1112         46.44 a           L1091         45.39 b           L1093         44.84 b           L1206         43.26 c           Okapi         45.01 b			L1206	34.72 <sup>i</sup>
L1112 46.44 a L1091 45.39 b L1093 44.84 b L1206 43.26 c Okapi 45.01 b	2016 2017		Okapi	36.33 <sup>h</sup>
Drought stress         L1091         45.39 b           L1093         44.84 b           L1206         43.26 c           Okapi         45.01 b	2010-2017		L1112	46.44 <sup>a</sup>
Drought stress         L1093         44.84 b           L1206         43.26 c           Okapi         45.01 b			L1091	45.39 <sup>b</sup>
L1206 43.26 ° Okapi 45.01 <sup>b</sup>		Drought stress	L1093	44.84 <sup>b</sup>
Okapi 45.01 <sup>b</sup>			L1206	43.26 °
			Okapi	45.01 <sup>b</sup>

Means in a column and a treatment followed by the same letter are not significantly different at 5% level.

The main effects of the sowing date and irrigation treatments were significant for CC and there was also a significant difference between the lines (p < 0.01). The CC was also affected by the interaction of the sowing date and irrigation (0.01 ) (Table 1).

A comparison of the mean interactions between the sowing date and irrigation indicated that the amount of CC decreased for the delayed sowing date of November 1 (0.87 mg g<sup>-1</sup> leaf fresh weight) as the plant hibernates with a weaker rosette, this disrupts the activity of the chloroplasts and CC and finally reduced photosynthetic activity. Also, drought stress causes chloroplast degradation and CC reduction by irrigation interruption from the silique formation stage to the next stage (Fig. 3). A study concerning rapeseed suggested that drought stress could influence chlorophylls a and b, as well as the total CC (Din *et al.*, 2011).

The interaction of the sowing date and irrigation showed that delayed sowing caused a significant decrease in the CC in both irrigation treatments, whereas drought stresses decreased CC significantly (Fig. 4). Photosynthetic pigments have a vital task in light absorbance and in the



**Fig. 3.** Interaction of sowing date and irrigation on chlrophyll content (CC) of the studied lines (2015-2017). Means with the same letter are not significantly different.



**Fig. 4.** Interaction of sowing date and irrigation on grain yield of the studied lines (2015-201 7). Means with the same letter are not significantly different.

production of reduction power (energy) in plants, they comprise a vast range of different pigments, which directly or indirectly play a role in photosynthesis (Jaleel et al., 2009). The total chlorophyll index in leaves is an effective index used to quantify photosynthetic potential and overall plant power (Hara et al., 2013). A decrease in CC was reported to be an indicator of plant reaction to drought stress (Bayat et al., 2009). Akram et al. (2018) reported that drought stress caused a significant reduction in growth, chlorophyll pigments, LRWC, and soluble proteins, while it also resulted in significant increases in relative membrane permeability, proline, glycine, betaine, malondialdehyde, phenolic compounds, and in the activities of catalase and peroxidase enzymes. In this research, the first sowing date raised the total CC due to the increased vegetative growth period. On the other hand, the plants cultivated on October 12 had an improved initial growth period because of the simultaneous primary growth of rapeseed seedlings with an optimal growth temperature occurring in mid-October and early November.

Drought stress harmed the total CC in rapeseed lines and reduced the total CC with an impact on the related metabolism in chlorophyll synthesis. The assessment of the physiological properties of rapeseed in response to drought stress demonstrated that soil moisture insufficiency for vegetative growth, flowering, and the silique development stages gave rise to a noteworthy decline in CC in different rapeseed lines relative to the irrigation treatments (Din *et al.*, 2011).

The main effects of year, sowing date, irrigation, and line on seed yield were significant (p < 0.01). There was also a significant interaction between the sowing date and irrigation (0.01 (Table 1). A comparison of themean interactions between the sowing date and irrigation showed a decreasing seed yield as a result of delayed sowing and drought stress, with the lowest seed yield (2080 kg ha<sup>-1</sup>) occurring on the November 1 sowing date and irrigation interruption from the silique formation stage to the next stage. The highest seed yield (5164 kg ha<sup>-1</sup>) was obtained on the October 12 sowing date and normal irrigation (Fig. 4). It is noteworthy that delayed sowing had a more pronounced effect on seed yield than drought stress because seed yield was higher under normal irrigation and sowing date (October 12) than normal irrigation and a delayed sowing date (November 1), as shown in Fig. 4. Hammac et al. (2017) declared that seed yield is profoundly affected by irrigation interruption at the flowering stage of rapeseed genotypes. Drought stress causing a decline in photosynthesis likely led to the inappropriate accumulation of photosynthates for seed filling and eventually reduced seed yield.

Seed yield had significant positive correlations with *LRWC*, CC, and LPC, while it presented significant negative correlations with SR, CT, and CLS (p < 0.01) (Table 5). The closest correlation was observed between SR and seed yield, suggesting an elevated seed yield with decreasing SR, which led to increased stomatal opening, a higher CO<sub>2</sub> exchange, increased photosynthetic activity, and finally, to an elevated seed yield. Pasban Eslam *et al.* (2017) studied the correlation between seed yield and physiological traits and reported that rapeseed genotypes with a higher *LRWC* and lower stomatal resistance had relatively higher seed yields under drought stress at the seed filling stage.

Stepwise regression was used to eliminate the effects of ineffective or less effective traits on seed yield in the regression model. Among the studied physiological traits, stomatal resistance, canopy temperature, and leaf proline were entered into Eq. (3), respectively (Table 6).

$$Y = 11041 - 136.7 X_2 - 129.9 X_3 - 57.53 X_5.$$
(3)

Y is seed yield, and  $X_2$ ,  $X_3$ , and  $X_5$  denote stomatal resistance, canopy temperature, and proline content, respectively. The model had an R<sup>2</sup> of 0.810, meaning that these traits account for more than 81% of seed yield variations. Stomatal resistance alone accounts for 79% of seed yield variations (Table 6). Stomatal resistance was introduced earlier in the model than the other traits and had the closest correlation (-0.89) with seed yield (Table 5). The addition

	SV	IRWC	SR	СТ	00	I PC	CLS
	1	LAWC	SK	CI		LIC	CLS
5 Y	1						
LRWC	0.783 *	1					
SR	-0.892 *	-0.915 *	1				
СТ	-0.889 *	-0.887 *	0.972 *	1			
CC	0.854 *	0.885 *	-0.957 *	-0.958 *	1		
LPC	0.754 *	0.984 *	-0.890 *	-0.858 *	0.859 *	1	
CLS	-0.885 *	-0.859 *	0.984 *	0.974 *	-0.967 *	-0.829 *	1

**Table 5.** Pearson correlation coefficients for seed yield, leaf relative water content (*LRWC*), stomatal resistance (SR), canopy temperature (CT), chlorophyll content (CC), leaf proline content (LPC), and carbohydrate of leaf solution (CLS) of the studied lines

Significant at p < 0.01.

**Table 6.** Stepwise regression for seed yield (dependent variable)

 and the other traits (independent variable)

	Stepwise regression stages				
Added trait to model	1	2	3		
Intercept	7109	9635	11041		
SR	-180.7	-99.83	-136.7		
СТ		-138.2	-129.9		
LPC			-57.53		
R <sup>2</sup>	0.795	0.804	0.811		

**Table 7.** Path analysis showing direct and indirect effects on the seed yield in rapeseed promising lines

	SR	СТ	LPC		
SR	<u>-0.675</u>	-0.376	0.157		
СТ	-0.656	<u>-0.386</u>	0.152		
LPC	0.600	0.330	<u>-0.178</u>		
Residual effect = $0.435$					

Underlined values have direct effects.

of other variables to the model did not significantly influence any further increase in  $\mathbb{R}^2$ . CT accounts for the highest grain yield variations in 30 genotypes of bread wheat under mild and intense drought stress conditions among such traits as 1000-grain weight, plant height, number of grains per panicle, and CC (Naroui Rad *et al.*, 2010).

SR had the greatest direct and negative effect (-0.675) on seed yield (Table 7). This trait had the most pronounced indirect effect on seed yield through CT (-0.376) and LPC (-0.157), respectively, and LPC exerted the least direct effect (-0.178) and the greatest indirect effect (0.930) on seed yield. It seems that high SR in the studied lines decreased CO<sub>2</sub> uptake and consequently decreased the photosynthetic rate, which ultimately lowered the seed yield. Therefore, the lines with a lower SR had a higher CO<sub>2</sub> uptake, photosynthetic rate, and yield (Taiz *et al.*, 2018). Since there was a significant positive correlation between

SR and CT, it appears that the transpiration rate decreased with increasing SR, followed by an increased CT in the studied lines.

## CONCLUSIONS

1. Delayed sowing and late-season drought stress led to significant increases in stomatal resistance, canopy temperature, and carbohydrate of leaf solution, but significant decreases in leaf relative water content, chlorophyll content, and seed yield in the studied lines. By evaluating traits such as stomatal resistance, canopy temperature, leaf relative water content, and chlorophyll content, it is likely that stress, increased stomatal resistance, and stomatal closure led to reduced leaf relative water content, chlorophyll content, CO<sub>2</sub> entrance and thus the nutritional processes diminished notably in the plant, which finally reduced the seed yield. Among the studied lines, the highest leaf relative water content, chlorophyll content, and seed yield belonged to L1206 whereas L1112 contained the highest carbohydrate of leaf solution level. Thus, L1112 was able to increase carbohydrate of leaf solution in the face of adverse conditions, resulting in the loss of crop yield and greater seed yield loss in the plant, which indicates the greater susceptibility of the line to the experimental conditions.

2. The amount of leaf proline content did not increase significantly due to delayed sowing and late-season drought stress, thus indicating the following points: (1) Under stress conditions, rapeseed plants increased their resistance by increasing the amount of carbohydrate of leaf solution. (2) Since the crop needs to use amino acids and energy (glucose) to produce proline, and the final goal of rapeseed is to produce oil (seed and oil yield), the crop achieved resistance with the accumulation of carbohydrate of leaf solution instead of increasing its resistance to stress through the production of proline.

3. The results of the correlation and stepwise regression showed that among the physiological traits in this study, stomatal resistance justified most of seed yield variations, with the greatest direct and negative effect on seed yield according to the path analysis results. Therefore, the measurement of stomatal resistance can be used as a parameter for the selection of resistant genotypes under stress conditions.

4. L1206 achieved the highest leaf relative water content, chlorophyll content and seed yield and the lowest stomatal resistance under delayed sowing and late-season drought stress. L1206 may be introduced as a superior line in order to select a line that can be cultivated in rotation with autumn crops and present favourable resistance to drought stress with the lowest yield loss at the end of the growing season in arid and semi-arid regions.

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#### REFERENCES

- Akram N.A., Iqbal M., Muhammad A., Ashraf M., Al-Qurainy F., and Shafiq S., 2018. Aminolevulinic acid and nitric oxide regulate oxidative defense and secondary metabolisms in canola (*Brassica napus* L.) under drought stress. Protoplasma, 255, 163-174. https://doi:10.1007/ s00709-017-1140-x
- Araus J.L., Slafer G.A., Royo C., and Serret M.D., 2008. Breeding for yield potential and stress adaptation in cereals. Critical Reviews in Plant Sci., 27, 377-412. https://doi: 10.1080/07352680802467736
- Ardakani M.R., Pietsch G., Moghaddam A., Raza A., and Friedel J.K., 2009. Response of root properties to tripartite symbiosis between lucerne (*Medicago sativa* L.), rhizobia and mycorrhiza under dry organic farming conditions. Am. J. Agric. Biol. Sci., 4, 266-277. https://doi: 10.3844/ ajabssp.2009.266.277
- Arnon D.I., 1949. Determination of chlorophyll concentration in leaf tissues of plants. Plant Physiol, 24, 1-15. https://doi. org/10.1104/pp.24.1.1
- Ashkiani A., Sayfzadeh S., Shirani Rad A.H., Valadabadi A., and Hadidi Masouleh H., 2020. Effects of foliar zinc application on yield and oil quality of rapeseed genotypes under drought stress. J. Plant Nutr. https://doi.org/10.1080/ 01904167.2020.1739299
- Balota M., Payne W.A., Evett S.R., and Lazar M.D., 2007. Canopy temperature depression sampling to assess grain yield and genotypic differentiation in winter wheat. Crop Sci., 47, 1518-1529. https://doi: 10.2135/cropsci2006.06.0383
- Bates L.S., Waldren R.P., and Teare I.D., 1973. Rapid determination of free proline for water-stress studies. Plant Soil, 39, 205-207. https://doi.org/10.1007/BF00018060
- Bayat F., Mirlohi A., and Khodambashi M., 2009. Effects of endophytic fungi on some drought tolerance mechanisms of tall fescue in a hydroponics culture. Russ. J. Plant Physl., 56, 563-570. https://doi:10.1134/S1021443709040104

- Cardoso A.A., Randall J.M., and McAdam S.A., 2019. Hydraulics regulate stomatal responses to changes in leaf water status in the fern *Athyrium filix-femina*. Plant Physiol., 179, 533-543. https://doi.org/10.1104/pp.18.01412
- Cashin P., Mohaddes K., Raissi M., and Raissi M., 2014. The differential effects of oil demand and supply shocks on the global economy. Energy Econ., 44, 113-134. https://doi. org/10.1016/j.eneco.2014.03.014
- Chun S.C., Paramasivan M., and Chandrasekaran M., 2018. Proline accumulation influenced by osmotic stress in arbuscular mycorrhizal symbiotic plants. Front Microbiol., 9, 1-30. https://doi.org/10.3389/fmicb.2018.02525
- Davaran Hagh E., Mirshekari B., Ardakani M.R., Farahvash F., and Rejali F., 2016. Optimizing phosphorus use in sustainable maize cropping *via* mycorrhizal inoculation. J. Plant Nutr., 39, 1348-1359. https://doi.org/10.1080/0190 4167.2015.1086797
- Din J., Khan S.U., Ali I., and Gurmani A.R., 2011. Physiological and agronomic response of canola varieties to drought stress. J. Anim. Plant Sci., 21, 78-82.
- Drake P., Froend R., and Franks P., 2013. Smaller, faster stomata: scaling of stomatal size, rate of response, and stomatal conductance. J. Exp. Bot., 64, 495-505. https://doi. org/10.1093/jxb/ers347
- Dubois M., Gilles K.A., Hamilton J.K., Rebers P.A., and Smith F., 1956. Colorimetric method for determination of sugars and relatedsubstances. Anal. Chem., 28, 350-356. https:// doi.org/10.1021/ac60111a017
- Enjalbert J.N., Zheng S., Johnson J.J., Mullen J.L., Byrne P.F., and McKay J.K., 2013. *Brassicaceae* germplasm diversity for agronomic and seed quality traits under drought stress. Ind. Crops Prod., 47, 176-185. https://doi. org/10.1016/j.indcrop.2013.02.037
- FAO, **2018.** World food and agriculture Statistical pocketbook 2018. Rome, 255 pp. http://www.fao.org/publications/card/ en/c/CA1796EN/
- Ferrat I.L. and Loval C.J., 1999. Relation between relative water content, nitrogen pools, and growth of *P. vulgaris* and *P. acutifolius* during water deficit. Crop Sci., 39, 467-474. https://doi.org/10.2135/cropsci1999.0011183X003900020 0028x
- Gaspar T., Franck T., Bisbis B., Kevers C., Jouve L., Hausman J.F., and Dommes J., 2002. Concepts in plant stress physiology. Application to plant tissue cultures. J. Plant Growth Regul., 37, 263-285.

https://doi.org/10.1023/A:1020835304842

- Grewal H.S., 2010. Water uptake, water use efficiency, plant growth and ionic balance of wheat, barley, canola and chickpea plants on a sodic vertisol with variable subsoil NaCl salinity. Agric. Water Manag., 97, 148-156. https:// doi.org/10.1016/j.agwat.2009.092
- Hammac W.A., Maaz T., Koeing R.T., Burke I.C., and Pan W.L., 2017. Water and temperature stresses impact canola (*Brassica napus* L.) fatty acid, protein and yield over nitrogen and sulfur. J. Agr. Food Chem., 47, 67-78. https://doi: 10.1021/acs.jafc.7b02778
- Hara M., Harazaki A., and Tabata K., 2013. Administration of isothiocyanates enhances heat tolerance in *Arabidopsis thaliana*. Plant Growth Regul., 69, 71-77. https://doi. org/10.1007/s10725-012-9748-5

- Henry C., John G.P., Pan R., Bartlett M.K., Fletcher L.R., Scoffoni C., and Sack L., 2019. A stomatal safety-efficiency trade-off constrains responses to leaf dehydration. Nat. Commun., 10, 1-9. https://doi.org/10.1038/s41467-019-11006-1
- Hoseinzade H., Ardakani M.R., Shahdi A., Asadi Rahmani H., Noormohammadi G., and Miransari M., 2016. Rice (Oryza sativa L.) nutrient management using mycorrhizal fungi and endophytic Herbaspirillum seropedicae. J. Integr. Agr., 15, 1385-1394. https://doi.org/10.1016/S2095-3119 (15)61241-2
- Ilkaee M.N., Paknejad F., Zavareh M., Ardakani M.R., and Kashani A., 2011. Prediction model of leaf area in soybean (*Glycine max* L.). Am. J. Agric. Biol. Sci., 6, 110-113. https:// doi: 10.3844/ajabssp.2011.110.113
- Jaleel C.A., Manivannan P., Wahid A., Farooq M., Al-Juburi H.J., Somasundaram R., and Panneerselvam R., 2009. Drought stress in plants: a review on morphological characteristics and pigments composition. Int. J. Agric. Biol., 11, 100-105.
- Khan M.A., Ashraf M.Y., Mujtaba S.M., Shirazi M.U., Khan M.A., Shereen A., Mumtaz S., Siddiqui M.A., and Kaler G.M., 2010. Evaluation of high yielding canola type brassica genotypes/ mutants for drought tolerance using physiological indices as screening tool. Pak. J. Bot., 42, 3807-3816.
- Krouma A., 2010. Plant water relations and photosynthetic activity in three Tunisian chickpea (*Cicer arietinum* L.) genotypes subjected to drought. Turk. J. Agric. For., 34, 257-264. https://doi: 10.3906/tar-0904-1
- Naroui Rad M.R., Jaafar H.Z.E., Akbari Moghaddam H., Poodineh O., and Moayedi H., 2010. Relation between physiological and some agronomic characteristics in selected genotypes of wheat in drought stress condition. J. Food Agric. Environ., 8, 891-893. https://doi.org/10.1234/ 4.2010.3427
- Nasiri A., Samdaliri M., Rad A.S., Shahsavari N., Kale A.M., and Jabbari H., 2017. Effect of plant density on yield and physiological characteristics of six canola cultivars. J. Sci. Agric., 1, 249-253. https://doi: 10.25081/jsa.2017.v1.819
- Pace R. and Benincasa P., 2010. Effect of salinity and low osmotic potential on the germination and seedling growth of rapeseed cultivars with different stress tolerance. Ital. J. Agro., 5, 69-77. https://doi.org/10.4081/ija.2010.69

- Papanatsiou M., Petersen J., Henderson L., Wang Y., Christie J.M., and Blatt M.R., 2019. Optogenetic manipulation of stomatal kinetics improves carbon assimilation, water use, and growth. Sci., 363, 1456-1459. https://doi.org/10.1126/ science.aaw0046
- Pasban Eslam B., Monirifar H., and Sadeghi Bakhtavari A.R., 2017. Morpho-physiological response of rapeseed (*Brassica napus* L.) genotypes to drought stress. Crop Breed. J., 7, 49-56. https://doi.org/10.22092/cbj.2018.116331.1019
- Reynolds M.P., Nagarajan S., Razzaque M.A., and Ageeb O.A.A., 2016. Heat tolerance. In: Application of Physiology in Wheat Breeding (Eds M.P. Reynolds, J.I. Oritz-Monastrio, and A. McNab) Mexico, D.F., CIMMYT. PP: 124-136.
- Sanchez-Rodriguez E., Rubio-Wilhelmi M.M., Cervilla L.M., Blasco B., Rios J.J., Rosales M.A., Romero L., and Ruiz J.M., 2010. Genotypic differences in some physiological parameters symptomatic for oxidative stress under moderate drought in tomato plants. Plant Sci., 178, 30-40. https:// doi.org/10.1016/j.plantsci.2009.10.001
- Saranga Y., Menz M., Jiang C.X., Wright R.J., Yakir D., and Paterson A.H., 2001. Genomic dissection of genotype × environment interactions conferring adaptation of cotton to arid conditions. Genome Res., 11, 1988-1995. https://doi. org/10.1101/gr.157201
- Taiz L., Zeiger E., Moller I.M., and Murphy A., 2018. Plant Physiology and Development. Oxford University Press, UK.
- Trueba S., Pan R., Scoffoni C., John G.P., Davis S.D., and Sack L., 2019. Thresholds for leaf damage due to dehydration: declines of hydraulic function, stomatal conductance and cellular integrity precede those for photochemistry. New Phytol., 223, 134-149. https://doi.org/10.1111/nph.15779
- Wu W., Ma B.L., and Whalen J.K., 2018. Enhancing rapeseed tolerance to heat and drought stresses in a changing climate: perspectives for stress adaptation from root system architecture. Adv. Agron., 151, 87-157. https://doi.org/10.1016/ bs.agron.2018.05.002
- Yan D., Easwaran V., Chau V., Okamoto M., Ierullo M., and Kimura M., 2016. NIN-like protein 8 is a master regulator of nitrate-promoted seed germination in Arabidopsis. Nat. Commun., 7, 13179. https://doi.org/10.1038/ncomms13179
- Yuncai H. and Schmidhalter U., 2005. Drought and salinity: A comparison of the effects of drought and salinity. J. Plant. Nutr. Soil Sci., 168, 541-549. https://doi:10.1002/ jpln.200420516